グロリオーサとエビネにおける組織培養苗生産方法の確立と実用化*

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Establishment of the methods for micropropagation and practical cultivation of the propagated plantlets in *Gloriosa* (Colchicaceae) and *Calanthe* (Orchidaceae)*

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Summary

Tissue culture is very effective for the rapid and mass propagation of plants, but many species are still difficult to propagate by this technology. In this study, *Gloriosa* and *Calanthe*, which have been recognized to be inefficient or hard to propagate by both conventional and tissue culture methods, were subjected to the tissue culture studies to establish efficient and practical methods for propagation and acclimatization, and to confirm the possibilities for the occurrence of somaclonal variations and virus elimination.

In *Gloriosa rothschildiana*, procedures for the micropropagation and cultivation of microtubers to flowering were established in this study. Both meristem and young shoot base explants were cultured on MS medium supplemented with 4.0 mg l⁻¹ BA and 0.1 mg l⁻¹ NAA, which resulted in the production of 5.0 and 47.2 shoots per explant, respectively, after 150 days of culture. Shoot multiplication of 2-3 times was successfully achieved by subculturing the excised shoots onto the same medium. During the subcultures, microtubers were formed at the base of each shoot by prolonged intervals of the subculture on the same medium. Approx. 85% of the microtubers of over 1 g thus produced successfully sprouted in *ex vitro* condition by the thermal treatment of 8°C for 2 weeks, followed by 30°C for 13 weeks. These results suggest that low temperature (8°C) was effective for breaking the dormancy of buds and high temperature for promoting the growth of buds. Sprouted tubers grew normally after planting in soil, and 80% of over 5 g tubers collected after 2 cycles of cultivation successfully produced flowers. No appreciable change was observed in the morphology including flower characters of the micropropagated plants. These results suggest that the micropropagation method established in this study could be used as the practical propagation method in the commercial cultivation of *Gloriosa*.

In *Calanthe*, there have been no successful reports on propagation with tissue culture. By improving the sterilization method for initial culture of *Calanthe sieboldii*, microbial contamination was prevented at 6% or less in the meristems excised from November to May. In the liquid culture, survival rate of meristems in B5 medium was better than that in MS and 1/2MS medium. Survival rate of meristems was also affected by the strengths (1, 1/2, 1/4) of macro (-CaCl₂), micro (+CaCl₂ and Fe-EDTA) and organic elements of B5 medium, and the highest survival rate was obtained when the concentration of micro elements (+CaCl₂ and Fe-EDTA) was reduced to the 1/4 strength of standard B5 medium. In this modified B5 medium supplemented with 2 mg 1⁻¹ BA, mass of shoot primordia was produced from shoot meristem explants. Then protocorm-like bodies (PLBs) were obtained from the mass of shoot primordia after transplanting onto agar medium with the same composition. These PLBs were vigorously and rapidly propagated by culturing in the same liquid medium. After two months of culture in the liquid medium, the number and fresh weight of PLBs became about 11 and 10 times, respectively.

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In the next step, the optimum ionic composition of medium for proliferation of PLBs was investigated by arranging systematic variations with 12 different ionic composition media (Ichihashi 1992a) with the use of PLBs induced from shoot meristem tip culture of *Calanthe aristulifera* and *C. striata*. The growth of PLBs was inhibited by higher NH₄⁺ and Ca²⁺ ratio, but promoted by higher ratios of K⁺ and H₂PO₄⁻. Calculated optimum cationic composition for *C. aristulifera* and *C. striata* were NH₄⁺: K⁺: Ca²⁺: Mg²⁺ = 10.4: 70.0: 9.6: 10.0, and 10.9: 68.1: 11.0: 10.0, respectively, and optimum anionic compositions were NO₃⁻: H₂PO₄⁻: SO₄²⁻ =42.1: 43.7: 14.2, and 43.0: 42.9: 14.1, respectively. It noted that NO₃⁻ was about half and 12-hold strength of H₂PO₄⁻ in comparison with control B5 medium with 1/4 micro elements.

Somaclonal variations in flower characters were investigated on 100 micropropagated *C. striata* plants. No obvious variations in flower shape and color were observed except two plants, which had larger and thicker flowers than original plants. These two variants were revealed to be tetraploid by flow cytometric analysis on relative nuclear DNA contents.

緒 言

本研究の背景 - 植物組織培養による増殖

植物の増殖方法として、種子による有性繁殖と株分け、挿し木(挿し芽)・取り木など植物体の一部を利用した無性繁殖(栄養繁殖)がある。種子による繁殖は、大量かつ一斉に増殖させる方法として有効である。しかし、種子繁殖は自家受粉が可能な植物の自殖を繰り返した種子の場合、または F1 雑種による種子の場合は、かなり均一な形質を有するが、両親が異なる場合など一般にヘテロ性が強く、形質にばらつきが出る。

一方、無性繁殖は遺伝的に同一であり、均一な形質をもっている. そのため、多年生草本や樹木に於いては、品種の特性を維持するためには不可欠な手段となっている. この繁殖方法の一つに、植物の組織を無菌的に、容器内で増殖させる組織培養という方法があり、株分け等では増殖困難な植物や短期間に大量に増殖させたい植物に利用されている. Morel and Martin(1955)が、ジャガイモを材料とし、また Morel(1960)がシンビデュームを材料として組織培養への取り組みが始まった. この分野は、研究が進み、多くの植物が組織培養により増殖が可能となり、商業的に利用されているが、まだ培養法が確立されていない植物も多い.

花き類の中では、高価で増殖しにくいラン科植物において茎頂培養が始まり、ウイルスフリーと大量増殖の両面から組織培養が行われている。カーネーション、カスミソウ、デルフィニウム、アルストロメリア、スターチス、ユリ、ガーベラなどで培養技術により苗生産が行われている(大澤 1994)。ここでは栽培下では増殖しにくく、切り花として大量

に生産されているが、新品種開発後の大量増殖に課題のあるグロリオーサ(Gloriosa)と今後有望な花きと考えられるエビネ属(Calanthe)について、組織培養を利用した増殖方法を検討したので、その結果を報告する.

① グロリオーサ (Gloriosa)

グロリオーサは、イヌサフラン科の半つる性植物で、南アフリカと東南アジアに5~6種類が分布している(De Hertogh and Le Nard 1993, Erhardt et al. 2002). 地下に塊茎を持つ多年草である. インドでは薬用植物として利用されており、さらにコルヒチンを生産する植物としても利用されている. 花が美しく、輝くような色彩と大きく特異な形から、観賞用植物として鉢植え、切り花として人気があり、広く普及している.

従来は rothschildiana 種や superba 種, superba var. lutea 種などに分類されていたが、最近ではすべての種が superba 種とされている (二宮 2010). しかし、本報では研究時に導入した種名である rothschildiana 種及び superba var. lutea 種として扱った.

Gloriosa の一種である G. rothschildiana は特に花が大きく、切り花として大量に栽培され、流通している. 高知県では年間 554 万本の切り花が出荷された (中国四国農政局高知統計・情報センター2004). さらに高知県、愛知県などの産地では品種改良も熱心に行われ、花色も従来からの赤色と黄色の複色タイプだけではなく、ピンク系、オレンジ系の品種もあり(石井ら2009)、白色系の品種も開発されている、繁殖は種子による有性繁殖、塊茎の分